

## **The effect of some beverage extracts on intestinal iron absorption**

**F. A. El-Shobaki, Z. A. Saleh, and N. Saleh**

National Research Centre, Dokki, Cairo, Egypt

*Summary:* The effect of some beverage extracts namely anise, mint, caraway, cumin, tilia, liquorice, karkade and tea, on the absorption of iron was tested in tied-off intestinal segments of rats. The rate of intestinal iron absorption was calculated in terms of an absorption index. The tannin, phytic acid and ascorbic acid contents of these beverages were analysed.

The results show that anise, mint, caraway, cumin, tilia, liquorice, arranged in decreasing order of their effect, promoted the absorption of iron. Karkade did not exert an appreciable effect while tea inhibited absorption. The results are discussed in relation to the content of these beverages of tannins, phytic or ascorbic acids. It is recommended to offer these beverages to children and also to adults as a preventive agent to iron deficiency anemia. Also can be used for the preparation of bioavailable medicinal iron.

*Zusammenfassung:* Es wurde der Einfluß von einigen Getränkeauszügen wie Anis, Minze, Kümmel, Cumin, Tilia, Süßholz, Karkade und Tee auf die Eisenresorption im Darm von Ratten untersucht. Die Eisenresorptionsrate wurde im Verhältnis zum Resorptionsindex berechnet. Der Tanningehalt sowie der Gehalt an Phytin- und Ascorbinsäure dieser Getränke wurde ermittelt.

Die Untersuchungsergebnisse zeigen, daß Anis, Minze, Kümmel, Cumin, Tilia und Süßholz in der Reihenfolge ihrer Erwähnung eine abnehmende Wirkung auf die Zunahme der Eisenresorption haben. Karkade hat keinen Einfluß auf die Eisenresorption, während Tee sie verhindert. Die Ergebnisse werden in bezug zum Tanningehalt und zum Phytin- oder Ascorbinsäuregehalt dieser Getränke diskutiert. Es wird empfohlen, diese Getränke Kindern und Erwachsenen als vorbeugendes Mittel bei Eisenmangelanämie zu geben.

*Key words:* iron, absorption, beverages, anemia, anise, mint, caraway, cumin, tilia, liquorice, karkade, tea

*Schlüsselwörter:* Eisenresorption, Getränke, Anämie, Anis, Minze, Kümmel, Cumin, Tilia, Süßholz, Karkade, Tee

### **I Introduction**

In most cases the iron deficiency anemia is due to the poor bioavailability of the ingested iron. This is particularly true in countries where the diet is predominantly composed of cereal or legumes. Iron present in such food, although appears to satisfy the nutritional requirements, yet, its bioavailability is poor (1, 2).

A highly bioavailable iron salt is essential for successful combating of anemia, that spread among different sectors of the population not only in developing countries but also in developed ones (3).

Beverages, such as anise, caraway, cumin, karkade, liquorice, mint, tilia and tea are given to children in our country some times as worm drinks between meals, and also as a remedy from diseases such as diarrhea or common cold. Little informations are available concerning the influence of these beverages on the bioavailability of iron.

The aim of the present study is to show how these beverages affect intestinal absorption of iron, and whether they are useful as promoters to iron absorption.

### Material and methods

The experiments were done on Sprague-Dawley albino rats of body weight ranging from 95–140 g and comprising both sexes. The groups of rats comprised from 6–7 litters.

The solution of beverages were prepared as usually done at homes. Table 1 shows the method of preparation and quantity of ingredient used. The iron test solution was ( $\text{FeCl}_3 \cdot 6 \text{H}_2\text{O}$ ) dissolved in 0.01 N HCl saline.

The phytate content in the beverage extract was determined by the method of Haug and Lantzsch (4). Tannin content was estimated by the method of Hagerman and Butler (5). Ascorbic acid was measured by the method of Association of Vitamin Chemists (6). The blood hemoglobin concentration was determined by the method of Betke and Savelsberg (7).

The dose given was formed by adding 1 ml from beverage extract to 1 ml of iron solution. This dose contains 500 n mol Fe, and was labelled with active iron ( $^{59}\text{Fe}$ ) obtained from (Institute National des Radioéléments, B-6220 Fleurus, Belgium).

Table 1. Method of preparation of beverage extract.

No.	Beverage	Means of preparation	g/100 ml
1	Anise	Seeds boiled with distilled water for 5 min	2.5 g/100 ml
2	Caraway	Seeds boiled with distilled water for 5 min	2.5 g/100 ml
3	Cumin	Seeds boiled with distilled water for 5 min	2.5 g/100 ml
4	Karkade	Seeds boiled with distilled water for 5 min	5 g/100 ml
5	Liquorice	Powder infused in distilled water for 3 h	5 g/100 ml
6	Mint	Dried leaves boiled with distilled water for 5 min	1.5 g/100 ml
7	Tea	Black tea added to boiling distilled water for 1 min	1.5 g/100 ml
8	Tilia	Dry leaves boiled with distilled water for 5 min	1.5 g/100 ml

Each dose contains an activity that measured about 100,000 cpm in the Gamma Scintillation Counter (Nuclear Interprise Scaler Ratemeter SR5). The pH of the dose was measured before injection.

The absorption experiments were done in tied off segments of the intestine in vivo (8). The segment measured 20 cm and comprised both the duodenum and the front part of the jejunum. After an overnight fast, the iron dose was injected into the tied-off segment during ether anaesthesia. One hour after dosing, blood was withdrawn by open-heart puncture. The intestinal segment was removed, washed three times, each with 20 ml of physiological saline. The liver was separated. The rest of the body was placed in a polyethylene bag.

The  $^{59}\text{Fe}$  activity in each of blood, liver, intestinal segment and the rest of the body was measured in the Gamma Counter. The measurements were corrected for the background and decay rate of the isotope.

The absorption index was calculated according to the following equation:

$$\text{Absorption index} = \frac{{}^{59}\text{Fe activity in blood} + \text{liver} + \text{rest of the body}}{{}^{59}\text{Fe activity of the given dose}} \times 100$$

## Results

The results of analysis of the beverages are shown in Table 2. The blood hemoglobin concentration of rats used in these studies ranged from 11.3–12.4 g/100 ml.

### *Absorption of iron*

The measured activity of  $^{59}\text{Fe}$  in the rest of the body, blood, liver, and intestine of rats given the iron dose with beverages from the family umbelliferae is shown in Table 3.

Compared to rats given iron alone the addition of each anise, caraway or cumin to the dose caused a significant increase in the  $^{59}\text{Fe}$  content of the measured organs. The intestinal segment contained significantly lower content of  $^{59}\text{Fe}$  due to addition of each of anise or cumin. The absorption index amounted to 14.1, 33.7, 30.0 and 30.9 % for each of iron alone, with anise, caraway or cumin respectively.

The  $^{59}\text{Fe}$  activity measured in the body of rats given other beverages with the iron is shown in Table 4.

Table 2. Ascorbic acid, tannin, phytic acid and iron contents of ingredients used in this study (mg/100 g dry weight).

No.	Beverage	Ascorbic acid	Tannin	Phytic acid	Iron
1	Anise	14.4	trace	trace	37.7
2	Caraway	7.2	trace	10.0	15.1
3	Cumin	7.2	trace	10.0	31.3
4	Karkade	36.0	trace	trace	128.5
5	Liquorice	10.8	trace	10.0	118.2
6	Mint	trace	133.0	trace	156.6
7	Tea	24.0	1733.0	28.0	31.2
8	Tilia	12.0	trace	8.0	24.7

Table 3. Mean values  $\pm$  S.E. of  $^{59}\text{Fe}$  activity in rest of the body, blood, liver and intestinal segment, 1 hour after injection of the labeled iron dose alone or with the added beverage solution in tied-off segments of normal rats.

Given dose (1 ml)		$^{59}\text{Fe}$ Activity (cpm) in			
		Rest of the body	Blood	Liver	Intestine
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$	Mean	10 470	2 424	1 161	77 378
	$\pm$ S.E.	1 565	434	146	6 129
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	27 030	4 653	2 038	52 049
	$\pm$ S.E.	5 976	518	434	7 771
Anise pH 2.3	P <	0.025	0.01	S.S.	0.05
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	22 804	5 242	1 970	70 869
	$\pm$ S.E.	3 752	840	285	6 666
Caraway pH 2.2	P <	0.025	0.025	0.05	N.S.
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	24 309	3 770	2 802	26 338
	$\pm$ S.E.	1 040	342	177	1 990
Cumin pH 2.4	P <	0.001	0.05	0.001	0.001

Anise, caraway, and cumin belong to the family umbelliferae.

Table 4. Mean values  $\pm$  S.E. of  $^{59}\text{Fe}$  activity in rest of the body, blood, liver and intestinal segment, 1 hour after injection of the labeled iron dose alone or with the added beverage solution in tied-off segments of normal rats.

Given dose (1 ml)		$^{59}\text{Fe}$ Activity (cpm) in			
		Rest of the body	Blood	Liver	Intestine
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ pH 2.1	Mean	10 470	2 424	1 161	77 378
	$\pm$ S.E.	1 565	434	146	6 129
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	12 347	2 011	837	46 409
	$\pm$ S.E.	2 009	143	94	6 362
Karkade pH 2.6	P <	N.S.	N.S.	N.S.	0.01
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	24 333	6 190	3 051	55 862
	$\pm$ S.E.	3 342	900	1 173	6 630
Mint pH 3.0	P <	0.005	0.005	N. S.	0.05
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	2 428	441	461	83 247
	$\pm$ S.E.	776	173	116	2 966
Tea pH 2.6	P <	0.001	0.005	0.005	N.S.
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	17 340	2 183	1 865	51 635
	$\pm$ S.E.	1 104	247	164	9 863
Tilia pH 2.4	P <	0.01	N.S.	0.01	0.05
$\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ +	Mean	15 989	2 531	1 731	23 762
	$\pm$ S.E.	898	223	175	4 522
Liquorice pH 3.1	P <	0.025	N.S.	0.05	0.001

The results show that mint and tilia enhanced absorption of iron with an absorption index of 33.6 % and 21.4 %. Karkade caused a non significant increase in the  $^{59}\text{Fe}$  content in the rest of the body (absorption index 15.2 %). Liquorice slightly enhanced the absorption of iron. The absorption index was 20.3 %. Tea markedly diminished absorption of iron (absorption index 3.3 %). The intestinal segment contained significantly lower content of  $^{59}\text{Fe}$  in case of karkade, mint, tilia and liquorice, but higher value in case of tea. However, such higher value is not significantly different from control.

## Discussion

In general, beverages that promoted the absorption of iron were anise, mint, caraway, cumin, tilia, liquorice, and karkade, while tea hindered iron absorption. Evaluation of the action of these beverages was based on the absorption index of each. The enhancement of iron absorption due to addition of each of these beverages to the iron dose depends on the net interaction of promoters or inhibitors of iron absorption present in the beverage. Anise extract is free of tannin and phytic acid. It also contains relatively higher content of ascorbic acid relative to caraway, cumin, and tilia. Tannin and some forms of phytic acid are known to diminish the absorption of iron (9, 10). The tannin content of tea is more than 20 times that present in some of these beverages. This may explain the inhibitory action of tea on iron absorption. In addition, it was found that phytic acid content of tea is the highest among other beverages analysed.

Although, caraway, cumin and liquorice contain relatively higher content of phytic acid yet, caraway and liquorice considerably enhanced iron absorption. It seems that this form of phytate is the monoferric phytate that was reported to exert no hindrance effect on iron absorption (11).

These three beverages do not contain tannins which may also contribute to their enhancing action on iron absorption.

Although the absorption index of iron in rats given caraway with the iron dose is less than that in case of anise, yet the  $^{59}\text{Fe}$  content of blood of rats given karaway with the iron dose is higher. This indicates that more iron is incorporated into red blood cells in rats given caraway, i.e. better utilization.

The other beverages namely mint, tilia, karkade and liquorice promoted the absorption of iron to varying degrees. A remarkable effect of mint is manifested in the marked increase of  $^{59}\text{Fe}$  content in blood and liver. This shows that in addition to the enhancing action of mint to iron absorption, it also enhances utilization and storage.

An important observation is that the intestinal  $^{59}\text{Fe}$  content is higher in case of beverages that hindered iron absorption, while it is lower in beverages that enhanced absorption. The accumulation of iron in the intestinal mucosa may be the cause of hindrance to iron absorption (8).

The promoting action of such beverages to iron absorption makes us recommend mothers to offer such drinks to their children from time to time or between meals. This will help to improve the bioavailability of iron in the gut. It is also possible to make use of such beverages for the preparation of medicinal iron.

*References*

1. Martinez-Torres C, Layrisse M (1974) Interest for study of dietary absorption and iron fortification. *World Rev Nutr Diet* 19:51
2. El-Shobaki FA, Saleh N (1986) The effect of food stuffs commonly consumed in Egypt on iron absorption and utilization. *J Sci Food Agric* 37:64
3. WHO, Tech Report Series (1968) No 405, pp 1-37
4. Haug W, Lantzsch HJ (1983) Sensitive method for the rapid determination of phytate in cereal products. *J Sci Food Agric* 34:1423
5. Hagerman AF, Butler TC (1978) Protein precipitation method for the quantitative determination of tannins. *J Agric Food Chem* 26:809
6. Association of vitamin chemists (1951) *Methods of Vitamin Assay*. Interscience Publishers, Inc, New York
7. Betke K, Savelsberg W (1950) Stufenphotometrische Hämoglobinbestimmung mittels Cyanhämoglobin. *Z Biochem* 320:431
8. El-Shobaki FA, Rummel W (1977) Mucosal transferrin and ferritin factors in the regulation of iron absorption. *Res Exp Med* 171:243
9. Roy SN, Mukherjee S (1979) Influence of food tannins on certain aspects of iron metabolism: Part 1. Absorption and excretion in normal and anemic rats. *Indian J Biochem Biophys* 16:93
10. Gillooly M, Bothwell TH, Charlton RW, Torrance JD, Bezwoda WR, MacPhail AP, Derman DP (1984) Factors affecting the absorption of iron from cereals. *Br J Nutr* 51:37
11. Morris ER, Ellis R (1976) Isolation of monoferric phytate from wheat bran and its biological value as an iron source to the rat. *J Nutr* 106:753

Received September 16, 1990

**Authors' address:**

Dr. F. A. El-Shobaki, Food Science and Nutritional Department, National Research Center, Dokki, Cairo, Egypt